

## Enhancement of the Enantioselectivity in Lipase-Catalyzed Kinetic Resolutions of 3-Phenyl-2*H*-azirine-2-methanol by Lowering the Temperature to $-40\text{ }^{\circ}\text{C}$

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We report here an efficient preparation of (*S*)-(+)-phenyl-2*H*-azirine-2-methanol ((*S*)-(+)-**1**) and its acetate ((*R*)-(–)-**2**) by a lipase-catalyzed kinetic resolution carried out preferentially at  $-40\text{ }^{\circ}\text{C}$  in ether under unusual conditions for enzyme. We disclosed that a lipase from *Pseudomonas cepacia* (lipase PS) exerts its function at such a very low temperature to markedly enhance the enantioselectivity. In an enzymatic reaction, the enantioselectivity in the kinetic resolution is temperature dependent and obeys the following thermodynamic equation:<sup>1</sup>

$$\ln E = \Delta\Delta S^{\ddagger}/R - \Delta\Delta H^{\ddagger}/(RT) \quad (1)$$

Hence, lowering the temperature can increase the enantioselectivity so far as the reaction is carried out below the racemic temperature  $T_r$ .<sup>1</sup> However, no report on exemplification of the theory by the enzymatic reaction below  $0\text{ }^{\circ}\text{C}$  is available<sup>2</sup> so far, because an enzyme is generally believed not to work effectively at such low temperatures. We found that the lipase-catalyzed reaction obeys the equation (eq 1) ranging from  $+30$  to  $-50\text{ }^{\circ}\text{C}$ . Recently, the chemoenzymatic synthesis has attracted much attention because of the demand for an environmentally-acceptable and total-cost-effective synthetic method. In these aspects, the lipase-catalyzed kinetic resolution<sup>3</sup> has been widely utilized as a reliable and readily available method for the resolution of racemic alcohols and carboxylic acid esters. In order to increase the enantioselectivity, a variety of methods, *e.g.*, reaction in appropriate organic media,<sup>3e,4</sup> use of additive,<sup>5</sup> choice of acyl donor,<sup>6</sup> and so on,<sup>7</sup> have been invented, while new and readily available methods for synthetic organic

chemists are still being sought. One problem to be solved in the lipase-catalyzed resolution is low enantioselectivity for chiral primary alcohols, except the relatively successful case for *meso*-compounds,<sup>8</sup> where the methodologies devised for secondary alcohols are not readily applicable to primary ones.<sup>9</sup> We now propose that a simple cooling of the reaction system to  $-40\text{ }^{\circ}\text{C}$  can be effective for enhancement of the enantioselectivity of the enzymatic reaction.

Azirine **1**<sup>10,11</sup> adopted here seems to be a useful chiral building block because the highly strained C=N double bond accepts a variety of chemical transformations such as a reduction to aziridine or an introduction of appropriate nucleophiles diastereoselectively.<sup>11</sup> Quite recently, the asymmetric synthesis of azirine derivatives has been the focus of several groups,<sup>12</sup> because naturally-occurring antibiotics containing the skeleton have been found. This class of compounds involves (*S*)-azirinomycin<sup>13</sup> and (*R*)-(–)-dysidazirine,<sup>14</sup> the latter of which has been recently synthesized by developing an asymmetric synthetic method of the azirine skeleton.<sup>12b</sup> The lipase-catalyzed resolution of chiral azirine has not been reported so far.

Racemic azirine ( $\pm$ )-**1** was prepared from cinnamyl alcohol by the reported method<sup>15</sup> with some modifications in 50% overall yield as shown in Scheme 1, which involves bromination of cinnamyl alcohol, reaction with  $\text{NaN}_3$  in DMSO, and subsequent dehydrobromination and then thermolysis to ( $\pm$ )-**1**. In the final thermocyclization step, the reaction temperature should be carefully controlled not to exceed  $100\text{ }^{\circ}\text{C}$  until the evolution of nitrogen ceases. Chromatographic purification gave pale yellow crystals (mp  $57\text{--}58\text{ }^{\circ}\text{C}$ ), which are stable enough to be stored in a refrigerator.

To begin, the conditions of the lipase-catalyzed transesterification of ( $\pm$ )-**1** were optimized according to the conventional method (Scheme 2). Thus, lipase PS was found to be a suitable lipase after screening commercially available lipases.<sup>16</sup> The reaction with an equimolar amount of vinyl acetate in diisopropyl ether at  $30\text{ }^{\circ}\text{C}$  was

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(1) Review: Phillips, R. S. *Trends Biotechnol.* **1996**, *14*, 13 and references cited therein.  $T_r$  is defined as  $\Delta\Delta H^{\ddagger}/\Delta\Delta S^{\ddagger}$ , a temperature at which there is no enantiomeric discrimination.

(2) (a) Optimization of the selectivity in a PLE-catalyzed hydrolysis in aqueous methanol at  $-10\text{ }^{\circ}\text{C}$ : Lam, L. K. P.; Hui, R. A. H. F.; Jones, J. B. *J. Org. Chem.* **1986**, *51*, 2047. No theoretical discussion on the temperature effect was made. (b) Increasing the temperature for high enantioselectivity: Yasufuku, Y.; Ueji, S. *Biotechnol. Lett.* **1995**, *17*, 1311.

(3) For example: (a) Jones, J. B. *Tetrahedron* **1986**, *42*, 3351. (b) Chen, C.-S.; Sih, C. J. *Angew. Chem., Int. Ed. Engl.* **1989**, *28*, 695. (c) Klivanov, A. M. *Acc. Chem. Res.* **1990**, *23*, 114. (d) Wong, C.-H.; Whitesides, G. M. *Enzymes in Synthetic Organic Chemistry*; Pergamon: Oxford, 1994. (e) *Enzyme Catalysis in Organic Synthesis*; Drauz, K., Waldmann, H., Eds.; VCH: New York, 1994; Vol. 1. (f) Faber, K. *Biotransformations in Organic Chemistry*; Springer-Verlag: Berlin, 1995. (g) *Enzymatic Reactions in Organic Media*; Koskinen, A. M. P., Klivanov, A. M., Eds.; Blackie Academic: Glasgow, 1996.

(4) For recent papers: (a) Ke, T.; Wescott, C. R.; Klivanov, A. M. *J. Am. Chem. Soc.* **1996**, *118*, 3366. (b) Nakamura, K.; Kinoshita, M.; Ohno, A. *Tetrahedron* **1995**, *51*, 8799.

(5) (a) Itoh, T.; Takagi, Y.; Murakami, T.; Hiyama, Y.; Tsukube, H. *J. Org. Chem.* **1996**, *61*, 2158. (b) Gao, Z.-W.; Sih, C. J. *J. Am. Chem. Soc.* **1989**, *111*, 6836.

(6) Ema, T.; Maeno, S.; Takaya, Y.; Sakai, T.; Utaka, M. *J. Org. Chem.* **1996**, *61*, 8610.

(7) (a) Pressure effect: Kamat, S. V.; Beckman, E. J.; Russell, A. J. *J. Am. Chem. Soc.* **1993**, *115*, 8845. (b) Immobilization on Florisil: Yamada, H.; Sugai, T.; Ohta, H.; Yoshikawa, S. *Agric. Biol. Chem.* **1990**, *54*, 1579.

(8) For example: (a) Yokomatsu, T.; Sato, M.; Shibuya, S. *Tetrahedron: Asymmetry* **1996**, *7*, 2743. (b) Hirose, Y.; Kariya, K.; Sasaki, I.; Kurono, Y.; Ebiike, H.; Achiwa, K. *Tetrahedron Lett.* **1992**, *33*, 7157. (c) Fuji, K.; Kawabata, T.; Kiryu, Y.; Sugiura, Y. *Tetrahedron Lett.* **1990**, *31*, 6663.

(9) Weissfloh, A. N. E.; Kazlauskas, R. J. *J. Org. Chem.* **1995**, *60*, 6959.

(10) Chemical resolution with low ee by using brucine: Stegman, W.; Uebelhart, P.; Heimgartner, H.; Schmid, H. *Tetrahedron Lett.* **1978**, 3091. The absolute configuration is not given.

(11) Review: Padwa, A.; Woolhouse, A. D. In *Comprehensive Heterocyclic Chemistry*; Lowowski, W., Ed.; Pergamon Press: New York, 1984; Vol. 7, Chapter 5, p 47.

(12) (a) Bucher, C. B.; Linden, A.; Heimgartner, H. *Helv. Chim. Acta* **1995**, *78*, 935. (b) Davis, F. A.; Reddy, G. V.; Liu, H. *J. Am. Chem. Soc.* **1995**, *117*, 3651. (c) Gentilucci, L.; Grijzen, Y.; Thijs, L.; Zwanenburg, B. *Tetrahedron Lett.* **1995**, *36*, 4665. (d) Verstappen, M. M. H.; Ariaans, G. J. A.; Zwanenburg, B. *J. Am. Chem. Soc.* **1996**, *118*, 8491.

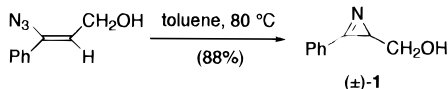
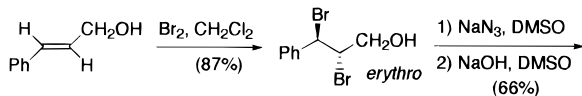
(13) (a) Stapley, E. O.; Hendlin, D.; Jackson, M.; Miller, A. K. *J. Antibiot.* **1971**, *24*, 42. (b) Miller, T. W.; Tristram, E. W.; Wolf, F. J. *J. Antibiot.* **1971**, *24*, 48.

(14) (a) Molinski, T. F.; Ireland, C. M. *J. Org. Chem.* **1988**, *53*, 2103. (b) Salomon, C. E.; Williams, D. H.; Faulkner, D. J. *J. Nat. Prod.* **1995**, *58*, 1463.

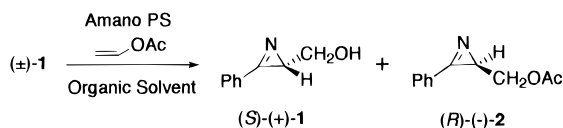
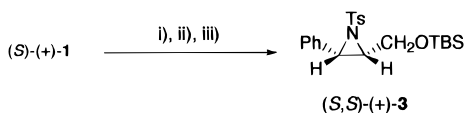
(15) (a) Hortmann, A. G.; Robertson, D. A.; Gillard, B. K. *J. Org. Chem.* **1972**, *37*, 322. (b) Padwa, A.; Rasmussen, J. K.; Tremper, A. *J. Am. Chem. Soc.* **1976**, *98*, 2605.

(16) Lipases showing the *E* values  $> 3$  (origin, *E* value, fast-reacting enantiomer): CHIRAZYME L-2 (*Candida antarctica*, 8, *R*), CHIRAZYME L-7 (porcine pancreas, 5, *R*), lipase AK (*Pseudomonas fluorescens*, 4, *R*).

Scheme 1



Scheme 2

Scheme 3<sup>a</sup>

<sup>a</sup> Key: (i) TBS-Cl, imidazole, THF (34%); (ii) LiAlH<sub>4</sub>, Et<sub>2</sub>O (51%); (iii) TsCl, Et<sub>3</sub>N, CHCl<sub>3</sub> (58%).

found to reach to a moderate conversion within a few hours, giving the *E* value<sup>17</sup> of 11 at best. Next, screening of the solvents<sup>18</sup> revealed that ether is the best choice, raising the *E* value to 17. In addition, vinyl acetate proved to give the highest selectivity among the several types of the acylating agents<sup>6</sup> examined. In spite of these efforts, the *E* value still remained around 17 and no other conventional means seemed to be applicable. The absolute configuration of (+)-**1**<sup>10</sup> thus obtained was determined to be *S* by correlation to authentic *N*-tosyl aziridine (*S,S*)-(+)-**3**<sup>19</sup> as shown in Scheme 3.

Then, we examined the temperature effect on the enantioselectivity in the prospects mentioned above. The reaction of (±)-**1** was carried out with lipase PS and vinyl acetate in ether at the temperature ranging from 30 to -60 °C as shown in Table 1. As the reaction system was cooled, the reaction rate was decreased as expected (convn h<sup>-1</sup> lipase mg<sup>-1</sup>), and thus, the amount of lipase was increased from 50 mg to 100, 200, and then 1600 mg at -60 °C so as to finish the reaction within a moderate time. The amount of the lipase, however, has no significant influence on the *E* value as examined at 30, 0, and -40 °C, respectively. Noteworthily, as the temperature was lowered the *E* values were markedly increased and reached up to 99 (at -40 °C), sufficient for practical use. Further cooling to -60 °C, however, began to lose the efficiency, decreasing the *E* value to 64.<sup>20</sup> The results listed in Table 1 are plotted in Figure 1 to give a straight line as a function of ln *E* and 1/*T* ranging from 30 to -50 °C. The parameters, ΔΔ*H*<sup>‡</sup> and ΔΔ*S*<sup>‡</sup>, calculated according to the theoretical equation (eq 1) are -3.0 kcal mol<sup>-1</sup> and -4.3 cal deg<sup>-1</sup> mol<sup>-1</sup>, respectively, and the racemic temperature *T<sub>r</sub>* calculated is 425 °C.

(17) Chen, C.-S.; Fujimoto, Y.; Girdaukas, G.; Sih, C. J. *J. Am. Chem. Soc.* **1982**, *104*, 7294.

(18) Organic solvent (*E* value): THF (15), acetone (15), AcOEt (15), CH<sub>3</sub>CN (14), toluene (11), *i*-Pr<sub>2</sub>O (11), benzene (9), cyclohexane (9), *n*-hexane (7).

(19) Fujii, N.; Nakai, K.; Habashita, H.; Hotta, Y.; Tamamura, H.; Otaka, A.; Ibuka, T. *Chem. Pharm. Bull.* **1994**, *42*, 2241.

(20) When the temperature was decreased below -40 °C, the lipase might begin to lose the conformational flexibility essential for the catalytic activity.

Table 1. Temperature Modulation in the Lipase-Catalyzed Resolution<sup>a</sup>

entry	<i>T</i> (°C)	lipase (mg)	% yield <sup>b</sup> (% ee <sup>c</sup> )		time (h)	<i>c</i> <sup>d</sup>	×10 <sup>3</sup> c/h/lipase mg	<i>E</i> <sup>e</sup>
			( <i>S</i> )-(+)- <b>1</b>	( <i>R</i> )-(-)- <b>2</b>				
1	30	50	50 (59)	41 (80)	2.0	0.42	4.2	17
2	30	200	44 (76)	45 (77)	0.5	0.50	5.0	18
3	20	50	45 (90)	55 (72)	4.3	0.56	2.6	19
4	0	50	54 (65)	45 (88)	6.0	0.43	1.4	31
5	0	100	42 (83)	49 (85)	3.0	0.49	1.6	31
6	0	200	57 (68)	43 (87)	1.4	0.44	1.6	28
7	-10	200	50 (77)	43 (86)	4.0	0.47	0.59	37
8	-20	200	49 (78)	42 (90)	4.5	0.46	0.51	46
9	-30	200	60 (54)	37 (94)	7.5	0.37	0.25	54
10	-40	200	66 (39)	22 (97)	8.0	0.29	0.18	84
11	-40	400	62 (46)	31 (97)	4.3	0.32	0.19	99
12	-50	400	60 (62)	38 (96)	8.5	0.39	0.11	86
13	-60	1600	55 (61)	37 (94)	6.0	0.39	0.041	64

<sup>a</sup> Conditions: (±)-**1** (50 mg, 0.34 mmol), vinyl acetate (29 mg, 0.34 mmol), dry Et<sub>2</sub>O (5 mL). <sup>b</sup> Isolated yield. <sup>c</sup> Determined by HPLC analysis using a Chiralcel OB-H column for **1** and Chiralcel OD-H for **2**, respectively. <sup>d</sup> Conversion calculated from *c* = ee (**1**) / (ee (**1**) + ee (**2**)) according to ref 17. <sup>e</sup> Reference 17.

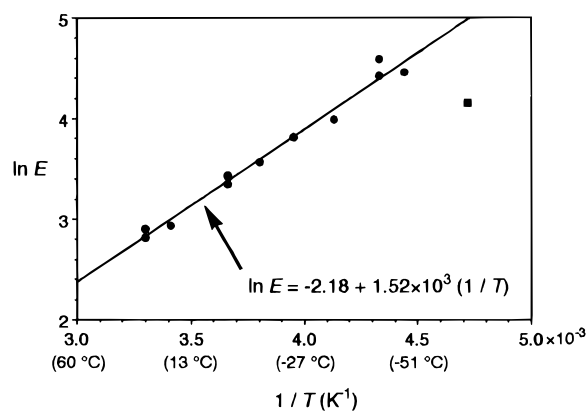


Figure 1. Temperature effect on the enantioselectivity.

These results suggest that the enantioselectivity of this lipase-catalyzed kinetic resolution is temperature-predictive even at such very low temperatures. Thus, lowering the temperature increases the *E* value, favoring the (*R*)-enantiomer in this case. The thermostability of the lipase in organic solvent<sup>3g,4</sup> enabled the reaction not only at higher temperatures<sup>21</sup> but also at such low temperatures. This temperature modulation will be a readily and generally applicable method to improve the enantioselectivity with theoretical prediction in the lipase-catalyzed resolution and should be a reliable method especially for primary alcohols. We are now actively investigating how to generalize the low-temperature modulating method by adopting to a variety of lipases and alcohols and how to utilize the chiral azirines (*S*)-(+)-**1** and (*R*)-(-)-**2** in organic synthesis.

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**Supporting Information Available:** Experimental procedures and spectral data for all compounds, including copies of <sup>1</sup>H and <sup>13</sup>C NMR spectra (18 pages).

JO970581J

(21) Lipase exhibits a high catalytic activity on heating at 100 °C: Zaks, A.; Klivanov, A. M. *Science* **1984**, *224*, 1249.